







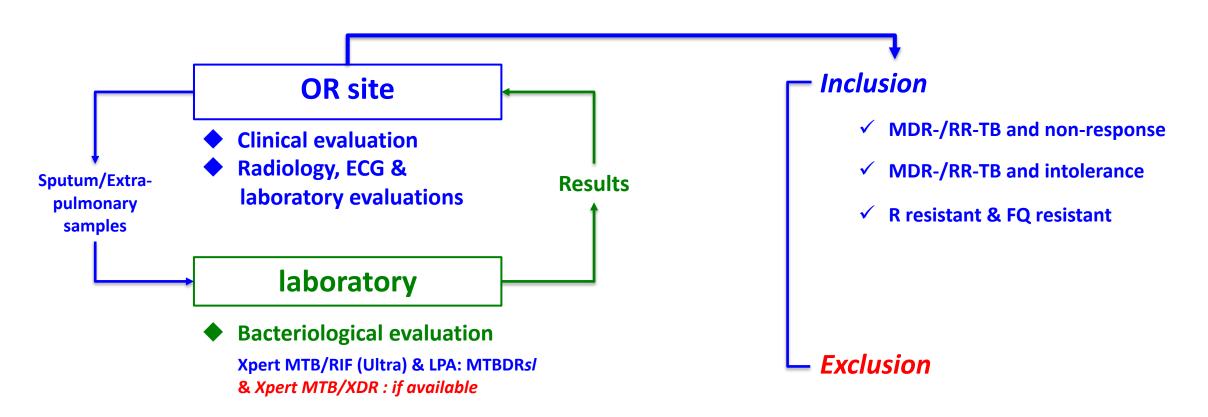
TOT Philippine: Bacteriological perspective







Patient inclusion: Bacteriological perspective





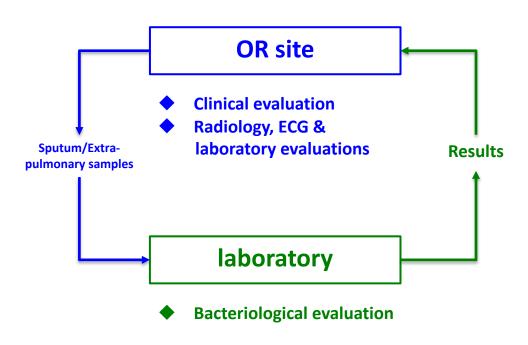








Baseline examinations in the laboratory



- Sputum smear
- Culture
- Sputum DST
 - Xpert MTB/RIF or Ultra
 - MTBDRs1
 - pDST for the second-line drugs

Isolates from positive cultures will be collected and stored for future research (pDST for BPaL, NGS, etc.).



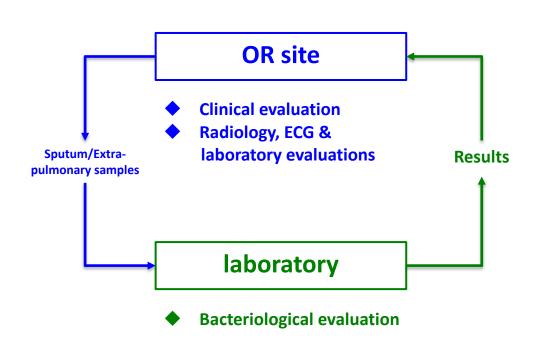








Examinations in monitoring evaluations



Extra-pulmonary samples (smear/culture/DST): if possible and no documented response to treatment

Monitoring will be done monthly, at the end of treatment, and at 6 and 12 months after treatment

- Sputum smear
- Culture
- pDST if smear or culture positive
 - at month 4, end of treatment or post-treatment follow up
 - can be done once pDST for BPaL is available.

Isolates from positive cultures will be collected and stored for future research (pDST for BPaL, NGS etc.).









New laboratory settings during LIFT-TB project

- Xpert MTB/XDR test platform
- pDST for the BPaL drugs: MGIT system

Further research

Nest generation sequencing (NGS): Comparison of genotypes and resistance conferring mutations in case of relapse suspects



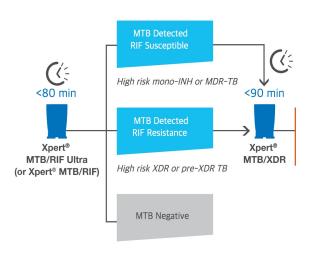






Xpert MTB/XDR

Can detect:
INH / FQs / SLIDs / ETH



5 machines (new 10 color detection model) and 4,000 cartridges will be ready to go soon in the Philippine

Even if XDR cartridge is available for the LIFT-TB project, do not replace LPA (MTBDRs/), but consider the Xpert XDR results as a reference: Even though WHO started to recommend to use Xpert XDR, accumulated clinical data up to now may not sufficient enough to replace MTBDRs/

Next generation sequencing

Hopely there will be no BPaL regimen failure cases, but if there are some, expect identify resistant conferring mutations for pretomanid

Table 6. Mutations associated with resistance to pretomanid in ≥ 2 isolates reported by ≥ 2 studies or ≥ 3 isolates reported by ≥ 1 study

| Gene | aa change [nt] | MIC summary (relative to parent for available isolates) | MIC change by mutant/study | Data summary | References |
|------|---------------------------|------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------|--------------------------|
| fbiC | V720I [G2158A] | ≥1-10× | ≥10× (1 isolate), ≥1× (0.36 mg/L selection concentration, 3 isolates, parent MIC ≤0.36 mg/L) ≥5× (1.8 mg/L selection concentration, 1 isolate, parent MIC ≤0.36 mg/L) | 5 in vitro selected mutants, MIC testing done for only 1 isolate | Haver 2015 ⁶⁰ |
| fbiC | P372S [C1114T] | ≥5× | \geq 5× (1.8 mg/L selection concentration, 3 isolates parent MIC \leq 0.36 mg/L) | 3 in vitro selected mutants, MIC testing not performed | Haver 2015 ⁶⁰ |
| fbiC | frameshift [ins C2549] | ≥5X | \geq 5× (1.8 mg/L selection concentration, 4 isolates, parent MIC \leq 0.36 mg/L) | 4 in vitro selected mutants, MIC testing not performed | Haver 2015 ⁶⁰ |
| ddn | S11* | ≥10× | ≥10× (15 isolates) | 15 in vitro selected mutants | Haver 2015 ⁶⁰ |
| ddn | Y133D | ≥5× | ≥5× (1.8 mg/L selection concentration, 3 isolates) | 3 in vitro selected mutants, MIC testing not performed | Haver 2015 ⁶⁰ |











THANK YOU

PLEASE CONTACT mlab.itrc@gmail.com FOR QUESITONS AND INQUIRIES

Acknowledgements: Jong Seok Lee, Hyejon Lee, Jinhee Lee, Jin-Kyung JUNG